

Journal Pre-proof

Protective effects of black seed (*Nigella sativa*) diet supplementation in common carp (*Cyprinus carpio*) against immune depression, oxidative stress and metabolism dysfunction induced by glyphosate

Morteza Yousefi, Hossein Adineh, Miriam Reverter, Mohammad Khademi Hamidi, Yury Anatolyevich Vatnikov, Evgeny Vladimirovich Kulikov, Seyed Hossein Hoseinifar, Hien Van Doan

PII: S1050-4648(20)30753-1

DOI: <https://doi.org/10.1016/j.fsi.2020.11.032>

Reference: YFSIM 7378

To appear in: *Fish and Shellfish Immunology*

Received Date: 2 October 2020

Revised Date: 26 November 2020

Accepted Date: 30 November 2020

Please cite this article as: Yousefi M, Adineh H, Reverter M, Hamidi MK, Vatnikov YA, Kulikov EV, Hoseinifar SH, Van Doan H, Protective effects of black seed (*Nigella sativa*) diet supplementation in common carp (*Cyprinus carpio*) against immune depression, oxidative stress and metabolism dysfunction induced by glyphosate, *Fish and Shellfish Immunology*, <https://doi.org/10.1016/j.fsi.2020.11.032>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2020 Published by Elsevier Ltd.



Seyed Hossein Hoseinifar and Hien Van Doan conceived and designed the experiments. Hossein Adineh and Mohammad Khademi Hamidi performed the experiments. Yury Anatolyevich Vatnikov and Evgeny Vladimirovich Kulikov analyzed the data. Miriam Reverter and Morteza Yousefi wrote and revised the paper. All authors read and approved the final manuscript.

Journal Pre-proof

1 Protective effects of black seed (*Nigella sativa*) diet supplementation in common carp (*Cyprinus*
2 *carpio*) against immune depression, oxidative stress and metabolism dysfunction induced by
3 glyphosate

4 Morteza Yousefi^a, Hossein Adineh^b, Miriam Reverter^c, Mohammad Khademi Hamidi^b, Yury
5 Anatolyevich Vatnikov^a, Evgeny Vladimirovich Kulikov^a, Seyed Hossein Hoseinifar^d, Hien Van
6 Doan^{e,f*}

7 ^a Department of Veterinary Medicine, Peoples' Friendship University of Russia (RUDN University), 6
8 Miklukho-Maklaya St, Moscow 117198, Russian Federation

9 ^b Department of Fisheries, Faculty of Agriculture and Natural Resources, Gonbad Kavous University, Gonbad
10 Kavous, Golestan, Iran

11 ^c Institute of Chemistry and Biology of the Marine Environment (ICBM), Carl von Ossietzky Universität
12 Oldenburg, Wilhelmshaven, Germany

13 ^d Department of Fisheries, Faculty of Fisheries and Environmental Sciences, Gorgan University of Agricultural
14 Sciences and Natural Resources, Gorgan, Iran

15 ^e Department of Animal and Aquatic Sciences, Faculty of Agriculture, Chiang Mai University, Chiang Mai
16 50200, Thailand

17 ^f Science and Technology Research Institute, Chiang Mai University, 239 Huay Keaw Rd., Suthep, Muang,
18 Chiang Mai 50200, Thailand

19 * Tel.: +66 90-029-9995; hien.d@cmu.ac.th

20

21 **Abstract**

22 Sustainable aquaculture arises as key to increase food production in the coming years. However, the
23 sector still faces many challenges such as the exposure of the cultured animals to pesticide-
24 contaminated water. Pesticides used in agriculture can reach aquaculture systems either directly
25 (integrated-agriculture aquaculture practices) or indirectly (soil leakage) and cause a broad range of

26 ecotoxicological effects on cultured fish and shellfish. Here, we studied how glyphosate affects
27 several haematological, biochemical, and immune parameters in common carp (*Cyprinus carpio*)
28 fingerlings, the fourth most important cultured fish species worldwide. We also evaluated the
29 potential of dietary supplementation with black seed (*Nigella sativa*, 0.25, 0.5 and 1%) to lower
30 glyphosate-associated toxicity. Our results showed that 14-day sub-lethal exposure of common carp
31 fingerlings to glyphosate increases oxidative stress, decreases antioxidant defences, affects several
32 metabolic pathways, and induced immune depression. Furthermore, we showed that fish fed with *N.*
33 *sativa*-enriched diets at 0.25, 0.5 and 1% for 60 days coped better with glyphosate exposure than
34 control fish and displayed more stable levels of biochemical serum parameters (total protein, albumin,
35 triglycerides, low-density lipoprotein LDL), cholesterol and high-density lipoprotein HDL), higher
36 levels of immune defences (lysozyme and immunoglobulin) and higher antioxidant enzymes
37 (superoxide dismutase SOD, glutathione peroxidase GPx) than control fish. Fish fed with all enriched
38 diets also displayed lower lipid peroxidation (malondialdehyde MDA), lower metabolic enzymes
39 (alanine aminotransferase ALT, aspartate aminotransferase AST and alkaline phosphatase ALP)
40 levels in blood serum and lower cortisol levels than control fish. Altogether, our results show that
41 dietary inclusion of black seed can be used as a sustainable bio-remediation strategy, mitigating many
42 of the negative effects of glyphosate exposure in fish.

43

44 Keywords: Glyphosate, *Nigella sativa*, *Cyprinus carpio*, Plant supplementation, Aquaculture

45

46 1. Introduction

47 As population continues to grow, one of the key societal challenges is to increase food production
48 while minimizing its impact in the environment [1]. The change in consumption patterns, including a
49 shift towards aquatic protein sources, especially from aquaculture, arises as a sustainable healthy
50 alternative to meat-based diets [2–4]. Aquaculture does not only provide a stable protein source that
51 could contribute to food security [5,6] but sustainable aquaculture practices require less natural

52 resources (i.e. water, space) than many terrestrial crops [3,7]. In fact, aquaculture is nowadays one of
53 the fastest-growing food-producing sectors, with estimates predicting a growth in aquaculture
54 production of at least 20% within the next ten years [8]. However, the intensification of aquaculture
55 practices to achieve higher productivities often entails decreases in water quality and increases in
56 disease outbreaks leading to high use of antimicrobials and emergence of antimicrobial resistance,
57 posing further health concerns [9,10]. The use of ecological intensification or integrated approaches
58 such as integrated agriculture-aquaculture (IAA) systems have been proposed as a way of increasing
59 aquaculture production, whilst increasing ecosystem resilience and therefore decreasing dependence
60 on antimicrobial drugs [11,12].

61 In IAA systems, fish are reared extensively (i.e. at low densities), using food resources mostly present
62 in the environment and optimising the use of water coming from surrounding agriculture systems
63 [13]. However, despite the many benefits of IAA, this can also result in a cross-contamination and
64 exposure of the cultured animals to agricultural residues such as pesticides [14,15]. Pesticides are
65 widely used in agriculture to control pests, however, once they reach the watercourses they often
66 display broad ecotoxicological effects on aquatic organisms [16,17]. Herbicides (i.e. pesticides used
67 for the control of deleterious weeds) account for nearly half of all the pesticides used worldwide, with
68 the highly controversial glyphosate N-(phosphonomethyl) being the most widely used herbicide for
69 weed control in many countries, including Iran [18]. Glyphosate, often applied as the commercial
70 formulation Roundup®, inhibits amino-acid synthesis in plants and is therefore widely used as a
71 broad-spectrum herbicide [19]. Several studies have shown deleterious effects of glyphosate on
72 aquatic species, ranging from genotoxicity [20], increase of oxidative stress [21], immune depression
73 [22], liver damage [23], and decrease in digestive functions and overall survival [24]. Furthermore,
74 environmentally relevant glyphosate concentrations were observed to increase the fish disease risk
75 [25], further indicating agriculture-related pesticides as a serious threat to fish production.

76 In order to ensure the aquaculture production growth needed to sustain the steady growing population
77 under such circumstances, effective strategies that mitigate pesticide toxicity in fish are needed. The
78 beneficial effects of fish dietary supplementation with medicinal plants on growth, haematology,

79 digestion, immunity, and disease resistance have been extensively studied during the last decade [26–
80 29]. More recently, studies started to show that plant enrichment can also improve fish resistance and
81 lower the oxidative stress caused by different stressors such as hypoxia [30], crowding stress [31,32]
82 or even disinfectant exposure such as copper sulphate [33]. To date, however, the potential of
83 medicinal plants or plant-derived compounds in protecting fish from pesticide-related toxicity remains
84 largely unexplored. Some of the few studies that have addressed the subject have shown that
85 supplementation of rainbow trout (*Oncorhynchus mykiss*) with *Ginkgo biloba* or *Vitis vinifera* extracts
86 mitigated the immune depressive effects associated with exposure to the organophosphate pesticide
87 diazinon [34,35], highlighting the potential of plant-enriched diets in lowering pesticide toxicity in
88 fish.

89 The common carp, *Cyprinus carpio*, is the fourth most important cultured fish species worldwide, and
90 is highly appreciated in many Middle-East countries such as Iran [36]. It is naturally found in the
91 Caspian Sea, where is extensively reared in most of the freshwater bodies including the farms in the
92 southern region of the Caspian Sea [37]. Many of these aquaculture industries are located near
93 agricultural farms and are therefore exposed to pesticide-contaminated water from the agricultural
94 drainage systems. In the present study, we aimed to investigate 1) how glyphosate affects several
95 haematological, biochemical, and immune parameters in common carp fingerlings and 2) the potential
96 of black seed (*Nigella sativa*) supplementation in mitigating glyphosate toxicity. Black seed or black
97 cumin is an herbaceous plant from the Ranunculaceae family that contains several bioactive
98 molecules such as thymoquinone, thymol, thymohydroquinone, dithymoquinone, p-cymene and
99 carvacrol [38]. Previous studies have shown that black seed supplemented diets enhanced immune
100 parameters including total protein, myeloperoxidase, bactericidal activity, immunoglobulin M and
101 lysozyme activity in rainbow trout, *Oncorhynchus mykiss* [39] and bactericidal activity and
102 phagocytic activity in common carp [26]. We now evaluate whether *N. sativa* might also display
103 beneficial effects when fish are exposed to pesticide-related stress.

104

105 2. Material and methods

106 2.1 Diet preparation

107 Black cumin (*Nigella sativa* L.) was purchased from a local shop (Gonbad Kavous, Iran), washed
 108 with deionized water, dried and powdered. A basal diet (Control) was formulated by mixing the
 109 feedstuffs, moistened with 300 ml water per kg (Table 1). The dough was pelleted via a meat grinder,
 110 and dried using a fan. Then, the resultant strings were crushed in an appropriate size. The
 111 experimental diets were prepared in the same way but adding 0.25, 0.5 and 1% black cumin powder
 112 (replaced with the wheatmeal). The feeds were kept at -20°C until use.

113 A basal diet (C) was formulated by mixing the feedstuffs, moistened with 300 ml water per kg (Table
 114 1). The dough was pelleted via a meat grinder, and dried using a fan. Then, the resultant strings were
 115 crushed in an appropriate size. The feeds were kept at -20°C until use. The experimental diets were
 116 prepared in the same way but adding 0.25, 0.5 and 1% black cumin powder (replaced with the
 117 wheatmeal). Proximate composition analysis of diets was performed according to the procedures of
 118 the AOAC (Association of Official Analytical Chemists) 2005.

119

120 Table 1. Ingredients and chemical composition of the basal diet.

Ingredients	Control	0.25	0.5	1.0
Fish meal	15	15	15	15
Meat meal	15	15	15	15
Soybean meal	23	23	23	23
Wheat meal	42	41.75	41.5	41
Fish oil	1	1	1	1
Soybean oil	1	1	1	1
Lysine ^a	0.5	0.5	0.5	0.5
Methionine ^b	0.5	0.5	0.5	0.5
Vitamin premix ^c	1	1	1	1
Mineral premix ^d	1	1	1	1
<i>Nigella sativa</i> powder	0	0.25	0.5	1.0

Dry matter	89.9	90.2	90.6	90.5
Crude protein	38.4	38.3	38.5	38.4
Crude lipid	8.69	8.70	8.76	8.95
Ash	7.09	7.12	7.09	7.03
Energy (MJ kg ⁻¹)	15.5	15.6	15.6	15.7

121 ^a Faravar Lysine Pars Co., Tehran, Iran.

122 ^B Mad Tiour Co., Sanandaj, Iran.

123 ^c The premix provided following amounts per kg of feed: A: 1000 IU; D3: 5000 IU; E: 20 mg; B5: 100 mg; B2:
124 20 mg; B6: 20 mg; B1: 20 mg; H: 1 mg; B9: 6 mg; B12: 1 mg; B4: 600 mg; C: 50 mg.

125 ^d The premix provided following amounts per kg of diet: Mg: 350 mg; Fe:13 mg; Co: 2.5 mg; Cu: 3 mg; Zn: 60
126 mg; NaCl: 3 g; dicalcium phosphate: 10 g.

127

128 2.2 Experimental procedure

129 The experiment was conducted at Gonbad Kavous University (Gonbad, Golestan, Iran), in accordance
130 with the ethics and animal care committee of Gonbad Kavous University. Common carp (*Cyprinus*
131 *carpio*) fingerlings were procured from a local farm and transported to the laboratory facilities in
132 Gonbad Kavous University. All fish were acclimatized for 14 days in 1,000 L tanks during which
133 were fed with the basal diet twice a day (2% of biomass). After the acclimation period, a total number
134 of 360 fish (mean weight 12.02 ± 0.29 g) were stocked in 12 fiberglass tanks (60 L) at a density of 30
135 fish per tank, to have triplicates for each of the four treatments. The fish were fed with the
136 abovementioned diets at 2.5% of biomass within 60 days. The biomass in each tank was weighed
137 biweekly to adjust the feed amount. The static water system with daily exchange was used in this
138 study. The tanks were continuously aerated, siphoned and 70% of the water was replaced daily. The
139 water physicochemical parameters including temperature (24.14 ± 0.61 °C), dissolved oxygen ($6.12 \pm$
140 0.58 mg L⁻¹), pH (7.19 ± 0.52), and total ammonia nitrogen (0.03 ± 0.011 mg L⁻¹) were measured
141 daily during the experimental period.

142 2.3 Exposure to glyphosate

143 The formulate product of glyphosate (N-phosphonomethyl glycine, 410 g glyphosate L⁻¹; Shanghai
144 Shenglian Chemical Co., Ltd.) was purchased from the distributor company (Iran). In order to expose
145 the fish to a glyphosate sub-lethal concentration, we first determined the glyphosate LC₅₀ value on
146 common carp fingerlings. Common carp fingerlings were separately exposed to 0.25, 0.5, 0.75, 1.5
147 and 2 mg L⁻¹ of glyphosate for 96 hours. Fish mortalities were recorded and the LC₅₀ value was
148 determined (0.489 mg L⁻¹) using the probit analysis [40].

149 After the 60-day feeding trial, the fish were exposed to glyphosate for 14 days by adding 0.122 mg L⁻¹
150 of glyphosate (25% of the LC₅₀ concentration) to each tank [35]. Glyphosate was added to each tank
151 after the daily water exchange (70%) in order to keep the concentration constant. To ensure that the
152 concentration of glyphosate is in the desired value, water samples (five times during the 14 days of
153 exposure) were taken from each tank and its concentration (0.122 ± 0.0011 mg/l) was measured using
154 GC-MS. During these two weeks, the fish were fed the same as the experimental period (4 different
155 treatments).

156 2.4 Growth performance and sampling

157 At the end of 60-day feeding trials, the fish were fasted for 24 hours and then their biomass was
158 recorded. Growth parameters and feed efficiency were calculated based on the fish initial weight
159 (IW), final weight (FW), Consumed feed (CI), and days of experiment (d) as follow [31]:

$$160 \text{ Specific growth rate (SGR, \% day}^{-1}\text{)} = 100 \times [(\ln \text{ FW} - \ln \text{ IW})/d]$$

$$161 \text{ Feed conversion ratio (FCR)} = \text{CI}/(\text{FW}-\text{IW})$$

$$162 \text{ Hepatosomatic index (HSI, \%)} = 100 \times [\text{liver weight (g)}/\text{whole body weight (g)}]$$

163 The fish blood was sampled (nine fish per treatment) for immunological and antioxidant assays at the
164 end of the 60-day feeding trial and after the 14-day exposure to glyphosate. The fish were gently
165 caught and placed in an anesthetic chamber (100 mg L⁻¹ eugenol) for 60 seconds. Blood was taken
166 from caudal vein and collected into sterile plastic tubes. The samples were left at 4 °C to clot, then
167 centrifuged for serum separation (1,200 g; 10 min).

168 After blood sampling, the fish were killed, disinfected in 0.1% benzalkonium chloride and the
169 intestine was sampled for analyzing the digestive enzyme activities. Fish were dissected and rinsed
170 with a sterile saline solution (0.85 % NaCl). The serum and intestine were frozen immediately in
171 liquid nitrogen and kept at -70°C for future analyses.

172 2.5 Biochemical and hematological analyses

173 Serum lysozyme activity was determined based on the sample ability to lyse *Micrococcus luteus* [41].
174 Briefly, *M. luteus* was suspended in phosphate buffered saline solution (pH 6.2), 1 mL of this
175 suspension was added to 30 μL of the serum samples and the decline on optical density (OD; at 530
176 nm) was recorded per minute within 3 min. The average decline on OD per min was calculated and
177 each 0.01-unit decline was considered as one unit of lysozyme activity. Alternative complement
178 activity (ACH_{50}) of the serum samples was assessed based on hemolytic activity of the samples as
179 previously described [42]. The target was sheep erythrocyte and reaction medium were veronal buffer
180 (pH 7) containing EGTA, gelatin and magnesium. The sheep erythrocyte was suspended in this
181 buffer. To this suspension, serial dilutions (10, 5, 2.5, 1.25, 0.625 and 0.312%) of the samples were
182 added and incubated at room temperature for 2 h. Then, stop solution (veronal buffer containing
183 EDTA and gelatin) was added and the mixture was centrifuged (1,000 g; 10 min). OD of the
184 supernatant was read at 412 nm. ACH_{50} activity was calculated according to Yano [43]. Total
185 immunoglobulin (Ig) was estimated by the method of Siwicki and Anderson [44]. Briefly, 100 μL
186 polyethylene glycol (12%) was added to equal volume of the serum sample and shaken for 2 h. Then
187 the mixture was centrifuged (1,000 g; 10 min) to precipitate Ig. Difference in protein content of the
188 samples before and after the centrifugation was equal to total Ig content.

189 SOD activity was determined based on Cytochrome C reduction rate using a commercial kit (Zellbio,
190 Berlin, Germany) as suggested by Hoseini et al. [45]. Serum catalase (CAT) activity was determined
191 based on decomposition rate of hydrogen peroxide according to Góth [46]. GPx activity was
192 measured based on conversion of glutathione to glutathione disulfide using a commercial kit (Zellbio,
193 Berlin, Germany) as suggested by Hoseini et al. [45]. MDA content was determined based on reaction
194 with thiobarbituric acid at 95°C for 1 h using a commercial kit (Zellbio, Hamburg, Germany) [45].

195 Glucose, total protein, albumin, cholesterol, triglycerides, HDL and LDL levels of serum were
196 evaluated spectrophotometrically using commercial kits (Pars Azmun Co. Ltd., Tehran, Iran) based on
197 previously described methods [47,48]. Serum level of cortisol was measured using the ELISA method
198 and a commercial kit (IBL, Gesellschaft für Immunchemie und Immunbiologie, Hamburg, Germany).

199 ALT, AST, and ALP activities were measured kinetically using Pars Azmun commercial kits (Pars
200 Azmun Co. Ltd., Tehran, Iran), as described previously [48].

201 Digestive tract samples were homogenized in 25 mM Tris-HCl buffer at pH 7.2, centrifuged (25,000 g
202 for 20 min) and the resultant supernatants collected. Activities of lipase, protease and amylase were
203 determined as described previously [49,50].

204 2.6 Statistical analysis

205 The data were checked for normality (Shapiro-Wilk test) and heterodascity (Levene's test) and
206 subjected to one-way ANOVA and Duncan test to investigate significant differences between the
207 different dietary treatments. Analysis of the glyphosate stress data was conducted using two-way
208 ANOVA (2 factors: glyphosate stress and black seed levels). SPSS v.22 was used for the analyses and
209 the results are presented as mean \pm SD.

210

211 3. Results

212 3.1. Fish parameters after 60 days *N. sativa* supplementation

213 3.1.1. Growth and feed assimilation

214 Common carp fingerlings fed diets supplemented with black seed (0.25, 0.5 and 1%) for 60 days
215 displayed higher ($P < 0.05$) final weight, weight gain (WG) and specific growth rate (SGR) than fish
216 fed with the control diet (Table 2). The feed conversion ratio (FCR) also improved significantly
217 (displayed lower values) in fish fed the plant-supplemented diets compared to control fish (Table 2).
218 Fish fed with enriched diets with 0.5 and 1% of *N. sativa* also presented significantly lower

219 hepatosomatic indices (HSI). Finally, no mortality was observed during the whole experiment in any
 220 of the treatments (Table 2).

221 Table 2. Effects of different dietary black cumin (*Nigella sativa* L.) powder levels on growth and feed
 222 performance of *Cyprinus carpio* after 60 days.

Treatments	Control	0.25	0.5	1.0
Initial weight (g)	12.00 ± 0.30	12.06 ± 0.32	11.85 ± 0.29	12.18 ± 0.27
Final weight (g)	36.10 ± 2.17 ^b	41.46 ± 2.06 ^a	40.63 ± 1.91 ^a	42.95 ± 2.51 ^a
WG (g)	24.10 ± 2.13 ^b	29.40 ± 1.86 ^a	28.78 ± 1.84 ^a	30.76 ± 2.35 ^a
SGR (% day ⁻¹)	1.83 ± 0.10 ^b	2.05 ± 0.06 ^a	2.05 ± 0.07 ^a	2.09 ± 0.08 ^a
FCR	1.91 ± 0.17 ^a	1.56 ± 0.09 ^b	1.60 ± 0.09 ^b	1.49 ± 0.11 ^b
HSI (%)	4.15 ± 0.32 ^a	3.90 ± 0.34 ^{ab}	3.67 ± 0.35 ^b	3.69 ± 0.26 ^b
Survival rate (%)	100	100	100	100

223 Values (mean ± SD for three replicate groups) in the same row not sharing a common superscript are
 224 significantly different ($P < 0.05$).

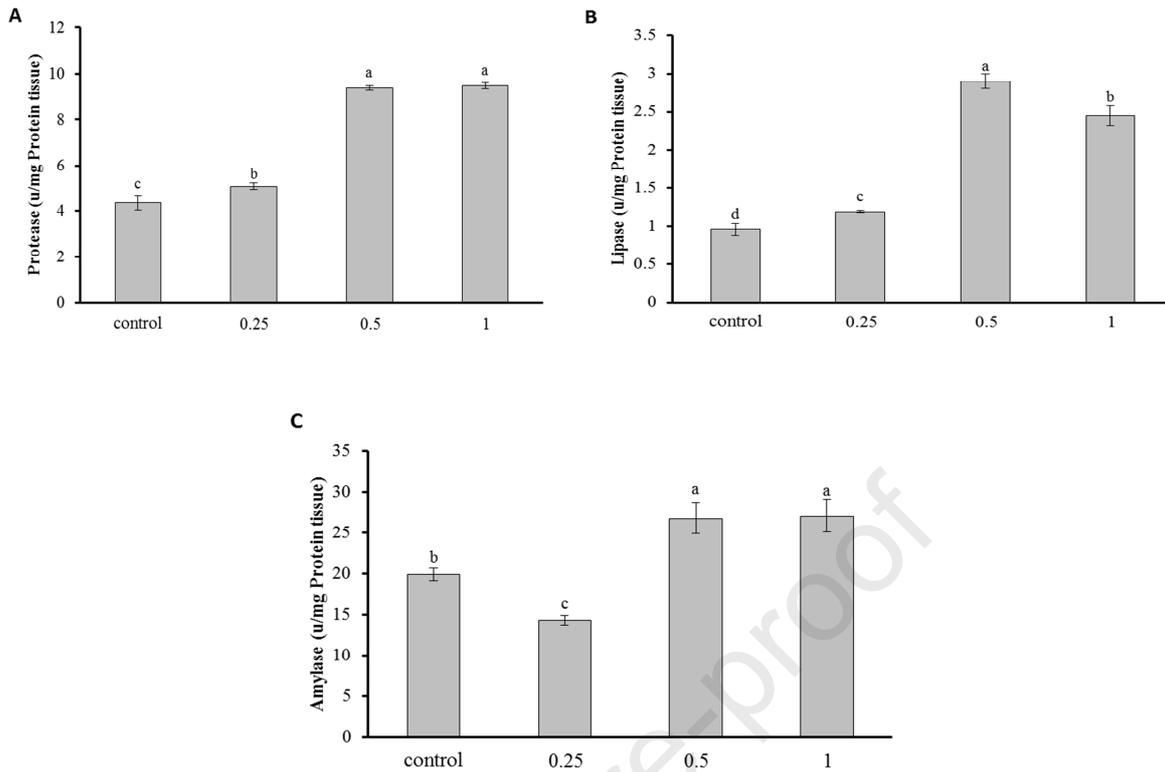
225 WG: weight gain, SGR: specific growth rate, FCR: feed conversion ratio, HSI: hepatosomatic index.

226

227 3.1.2. Digestive enzyme activities

228 The inclusion of black seed powder in *C. carpio* fingerlings diet enhanced the activity of digestive
 229 enzymes in most treatments. All fish fed with plant-enriched diets displayed significantly higher
 230 protease and lipase activities than control fish (Figure 1). The highest protease activities were found in
 231 fish fed with diets enriched containing 0.5 and 1% of black seed, whilst the highest lipase activity was
 232 found in fish fed with diets supplemented with 0.5% of *N. sativa* (Figure 1). Amylase activities were
 233 also significantly higher in fish fed with enriched diets at 0.5 and 1%; however, fish fed with diets
 234 enriched with 0.25% of black seed displayed significantly lower amylase levels than control (Figure
 235 1).

236



237

238

239

240 Fig. 1. Digestive enzymes activities (U mg^{-1} protein) of common carp fed diets with different levels of dietary
 241 *N. sativa* for 60 days. Different letters show significant difference among the treatments (Duncan test).

242

243 3.2. Fish parameters before and after glyphosate exposure

244 3.2.1. Antioxidant enzyme activities

245 MDA, which is an indicator of lipid peroxidation, increased significantly in fish exposed to
 246 glyphosate for 14 days, with fish fed with a control diet displaying the highest MDA levels and fish
 247 fed with an enriched diet (1% black seed), displaying the lowest levels (Table 3). Catalase levels also
 248 increased significantly in fish exposed to glyphosate (when all treatments combined), but no
 249 significant differences were observed in the levels of catalase amongst the treatments (Table 3). The
 250 dietary inclusion of black seed significantly enhanced the activity of SOD and GPx after exposure to
 251 glyphosate compared to a control diet. The highest SOD activities in fish exposed to glyphosate were
 252 observed in fish fed with a supplemented diet at 0.25 and 0.5% (Table 3). Moreover, the highest GPx

253 activity in fish exposed to glyphosate were observed in fish fed with a supplemented diet at 0.25%
 254 followed by 1% (Table 3).

255

256 Table 3: Serum SOD, GPx and CAT activities and MDA level in common carp fed with different levels of *N.*
 257 *sativa* powder for 60 days followed by 14 days exposure to glyphosate. Different lowercase letters within a
 258 column show significant effects of dietary *N. sativa* levels before or after glyphosate exposure. Different
 259 uppercase letters within a column show significant difference among the treatment combinations (interaction
 260 effects of *N. sativa* × glyphosate exposure). “De” shows significant decrease, while “In” shows significant
 261 increase in the tested parameters after the glyphosate exposure.

262

Experiment groups	parameters			
	Catalase (U/ml)	SOD (U/ml)	GPx (U/ml)	MDA (μmol/l)
Before Stress				
0	102.22 ± 8.95	32.96 ± 1.36 ^a CD	167.00 ± 5.65 ^{ab} DE	52.52 ± 5.01 ^a CD
0.25	101.51 ± 5.70	35.11 ± 1.71 ^a C	165.58 ± 3.14 ^b E	52.04 ± 4.29 ^a CD
0.5	108.13 ± 2.86	36.02 ± 1.25 ^a BC	173.27 ± 4.39 ^{ab} CD	44.35 ± 4.26 ^{ab} DE
1.0	105.33 ± 2.37	35.30 ± 2.59 ^a C	174.80 ± 2.26 ^a C	43.16 ± 3.75 ^b E
After Stress				
0	113.02 ± 5.33	31.80 ± 1.07 ^b D	162.50 ± 2.56 ^c E	69.83 ± 2.34 ^a A
0.25	110.16 ± 7.43	41.07 ± 0.92 ^a A	191.42 ± 3.15 ^a A	64.89 ± 5.13 ^a AB
0.5	108.88 ± 4.96	39.25 ± 2.52 ^a A	183.38 ± 4.64 ^b B	60.96 ± 4.71 ^a B
1.0	112.38 ± 4.25	38.41 ± 0.52 ^a AB	187.09 ± 4.30 ^{ab} AB	60.29 ± 6.09 ^a BC
Two-way ANOVA				
Stress	P = 0.009	P < 0.001	P < 0.001	P < 0.001
Diet	NS	P < 0.001	P < 0.001	NS
Interaction	NS	P = 0.015	P < 0.001	P = 0.006

263 Abbreviations: SOD: superoxide dismutase; GPx: glutathione peroxidase; MDA: malondialdehyde; De:
 264 decrease; NS: not significant.

265

266 3.2.2. Serum biochemical parameters in fish

267 Fish exposed to glyphosate for 14 days displayed significantly lower levels of serum total protein and
 268 albumin than naïve fish (i.e. not exposed to glyphosate) (Table 4). Within exposed individuals, fish
 269 fed with *N. sativa* enriched diets displayed significantly higher levels of total protein and albumin
 270 than fish fed with control diet. In fact, diet supplementation with black seed lowered the negative
 271 effect of glyphosate on total protein (21% total protein decrease in control fish vs. 8-13% in treated
 272 fish) and albumin (43% albumin decrease in control fish vs. 13-18% albumin decrease in treated fish)
 273 levels (Table 4). Serum total cholesterol and LDL levels were significantly higher in fish fed with *N.*
 274 *sativa* enriched diets compared to control (Table 4). Although both cholesterol and LDL decreased
 275 when fish were exposed to glyphosate, control fish and fish supplemented with the lowest level of
 276 black seed experienced much steeper decreases (21-38% decrease in cholesterol and 38% decrease in
 277 LDL) than fish fed with diets enriched in 0.5 and 1% of black seed (3-6% decrease in cholesterol and
 278 3-17% decrease in LDL) (Table 4). Before glyphosate exposure, the diet enriched in 0.5% of black
 279 seed displayed significantly higher HDL level than fish fed with control diet; however, after
 280 glyphosate exposure, all treatments showed significantly higher HDL level than fish fed with control
 281 diet. Moreover, after glyphosate exposure, HDL levels decreased significantly in control fish; whilst
 282 HDL did not change in fish that were fed diets supplemented with *N. sativa* (Table 4). Before
 283 exposure to glyphosate, the highest and lowest triglyceride levels were observed in fish fed with
 284 control and 1% group, respectively; whilst the highest and lowest triglyceride levels were observed in
 285 fish fed with 1% black seed supplemented diet and control group, respectively.

286 Table 4: Serum biochemical parameters in common carp fed with different levels of *N. sativa* powder for 60
 287 days followed by 14 days exposure to glyphosate. Different lowercase letters within a column show significant
 288 effects of dietary *N. sativa* levels before or after glyphosate exposure. Different uppercase letters within a
 289 column show significant difference among the treatment combinations (interaction effects of *N. sativa* ×
 290 glyphosate exposure). “De” shows significant decrease, while “In” shows significant increase in the tested
 291 parameters after the glyphosate exposure.

292

Experiment	Parameters					

groups	Total Protein (g/dL)	Albumin (g/dL)	Cholesterol (mg/dL)	Triglycerides (mg/dL)	LDL (mg/dL)	HDL (mg/dL)
Before Stress						
0	2.87 ± 0.19	1.80 ± 0.17A	147.94 ± 3.19 ^b C	184.52 ± 5.30 ^a A	73.32 ± 2.11 ^c D	41.03 ± 2.13 ^b C
0.25	2.98 ± 0.28	1.75 ± 0.24AB	153.53 ± 4.62 ^b C	129.51 ± 2.12 ^d D	85.06 ± 2.87 ^b B	45.74 ± 2.15 ^{ab} ABC
0.5	3.10 ± 0.11	1.77 ± 0.07AB	172.20 ± 5.23 ^a A	142.58 ± 4.60 ^c C	96.73 ± 2.84 ^a A	48.83 ± 4.73 ^a A
1.0	3.20 ± 0.17	1.88 ± 0.10A	173.44 ± 6.24 ^a A	157.83 ± 2.89 ^b B	98.07 ± 1.85 ^a A	44.12 ± 2.26 ^{ab} ABC
After Stress						
0	2.27 ± 0.06 ^b	1.03 ± 0.10 ^b D	92.20 ± 1.87 ^e E	109.06 ± 1.71 ^d E	45.21 ± 1.67 ^d F	32.73 ± 2.84 ^d D
0.25	2.73 ± 0.14 ^a	1.52 ± 0.11 ^a BC	121.47 ± 3.01 ^b D	139.71 ± 1.64 ^b C	51.57 ± 2.48 ^e E	42.67 ± 3.05 ^b BC
0.5	2.68 ± 0.07 ^a	1.46 ± 0.06 ^c C	167.37 ± 2.88 ^a AB	130.44 ± 2.26 ^d D	94.11 ± 1.16 ^a A	49.01 ± 4.42 ^a A
1.0	2.89 ± 0.20 ^a	1.62 ± 0.14 ^a ABC	162.75 ± 7.20 ^a B	181.20 ± 1.66 ^a A	80.80 ± 2.04 ^b C	47.14 ± 1.79 ^{ab} AB
Two-way ANOVA						
Stress	P < 0.001 De	P < 0.001 De	P < 0.001 De	P < 0.001 De	P < 0.001 De	NS
Diet	P = 0.002	P = 0.006	P < 0.001	P < 0.001	P < 0.001	P < 0.001
Interaction	NS	P = 0.011	P < 0.001	P < 0.001	P < 0.001	P = 0.034

293 Abbreviations: HDL: high-density lipoprotein cholesterol; LDL: low-density lipoprotein cholesterol; De:

294 decrease; NS: not significant.

295 The metabolic enzymes AST, ALT and ALP varied significantly in a dose-dependent manner in fish
 296 fed with different levels of *N. sativa*, with control fish displaying the highest levels and fish fed with
 297 the highest level of black seed (1%) displaying the lowest AST, ALT and ALP levels (Table 5).

298 Exposure of fish to glyphosate induced a significant increase in AST and ALT in all fish treatments,
 299 but levels remained the lowest in fish fed with the diet enriched with 1% of black seed (Table 5).

300 Moreover, after glyphosate exposure, the highest and lowest ALP level were observed in fish fed with
 301 control and 0.25% group, respectively.

302 Cortisol and glucose levels increased significantly in fish that were exposed to glyphosate for 14 days
 303 (Table 5). The inclusion of black seed in carps' diet resulted in a significant reduction of both serum
 304 cortisol and glucose levels, both in naïve fish and fish exposed to glyphosate (Table 5). The lowest
 305 values of cortisol in fish exposed to glyphosate were achieved at 1% black seed supplementation,

306 whereas the lowest glucose values in glyphosate-exposed fish were observed at 0.25%
307 supplementation (Table 5).

308 Table 5: Serum metabolic enzymes (AST, ALT and ALP), cortisol and glucose levels in common carp fed with
309 different levels of *N. sativa* powder for 60 days followed by 14 days exposure to glyphosate. Different
310 lowercase letters within a column show significant effects of dietary *N. sativa* levels before or after glyphosate
311 exposure. Different uppercase letters within a column show significant difference among the treatment
312 combinations (interaction effects of *N. sativa* × glyphosate exposure). “De” shows significant decrease, while
313 “In” shows significant increase in the tested parameters after the glyphosate exposure.

Experiment groups	Parameters				
	AST (u/L)	ALT (u/L)	ALP (u/L)	Cortisol (ng/mL)	Glucose (mg/dL)
Before Stress					
0	291.52 ± 9.21 ^a D	20.47 ± 1.29 ^a E	159.22 ± 1.88 ^a A	187.18 ± 8.47 ^a CD	106.72 ± 1.15 ^a AB
0.25	277.75 ± 7.37 ^b E	18.34 ± 0.73 ^b F	145.54 ± 3.42 ^b B	170.32 ± 10.33 ^{ab} DE	92.38 ± 2.99 ^b C
0.5	250.42 ± 6.10 ^c F	17.20 ± 0.34 ^c F	135.66 ± 3.12 ^c C	156.54 ± 17.67 ^b E	78.05 ± 1.31 ^c D
1.0	245.16 ± 6.03 ^c F	14.08 ± 1.10 ^c G	114.47 ± 3.18 ^d E	158.74 ± 5.12 ^b E	64.94 ± 2.88 ^d E
After Stress					
0	385.09 ± 7.19 ^a A	33.00 ± 1.02 ^a A	138.12 ± 2.58 ^a C	258.17 ± 5.27 ^a A	112.35 ± 2.87 ^a A
0.25	338.71 ± 2.05 ^b B	26.29 ± 0.73 ^c C	112.45 ± 1.28 ^e E	204.73 ± 6.24 ^{bc} BC	92.74 ± 7.51 ^b C
0.5	342.42 ± 6.32 ^b B	29.55 ± 1.05 ^b B	128.03 ± 3.16 ^b D	221.78 ± 14.38 ^b B	107.04 ± 1.84 ^a AB
1.0	325.16 ± 4.41 ^c C	24.60 ± 0.61 ^d D	125.50 ± 1.44 ^b D	197.30 ± 15.67 ^c C	105.02 ± 3.11 ^a B
Two-way ANOVA					
Stress	P < 0.001 In	P < 0.001 In	P < 0.001 De	P < 0.001 In	P < 0.001 In
Diet	P < 0.001	P < 0.001	P < 0.001	P < 0.001	P < 0.001
Interaction	P = 0.001	P = 0.002	P < 0.001	P = 0.028	P < 0.001

314 Abbreviations: ALT: alanine aminotransferase; AST: aspartate aminotransferase; ALP: alkaline phosphatase

315 3.2.3. Serum immune parameters

316 Levels of all three immune parameters measured in this study (ACH₅₀, lysozyme and total
317 immunoglobulin) were significantly reduced when carps were exposed to glyphosate for 14 days
318 (Table 6). Before glyphosate exposure, the highest lysozyme activity was observed in fish fed with a

319 diet supplemented with 1% black seed. Moreover, 0.5 and 1% black seed supplemented diet fish
 320 displayed the highest total immunoglobulin levels. After exposure to glyphosate, all treatments
 321 showed higher levels of lysozyme and total immunoglobulin than control group. ACH₅₀ levels did not
 322 vary significantly amongst different treatments (Table 6).

323 Table 6: Alternate complement activity (ACH₅₀), lysozyme activity and total immunoglobulin levels in common
 324 carp fed with different levels of *N. sativa* powder for 60 days followed by 14 days exposure to glyphosate.
 325 Different lowercase letters within a column show significant effects of dietary *N. sativa* levels before or after
 326 glyphosate exposure. Different uppercase letters within a column show significant difference among the
 327 treatment combinations (interaction effects of *N. sativa* × glyphosate exposure). “De” shows significant
 328 decrease, while “In” shows significant increase in the tested parameters after the glyphosate exposure.

	ACH50 (U/mL)	Lysozyme activity (u/mL/min)	Total Immunoglobulin (mg/mL)
Before Stress			
0	136.20 ± 3.71	32.40 ± 0.52 ^{bB}	16.96 ± .075 ^b
0.25	138.14 ± 2.92	33.00 ± 1.19 ^{bB}	17.79 ± 0.36 ^{ab}
0.5	139.31 ± 3.13	33.83 ± 1.36 ^{bB}	18.38 ± 0.70 ^a
1.0	137.66 ± 2.81	39.24 ± 1.44 ^{aA}	18.54 ± 0.81 ^a
After Stress			
0	129.00 ± 2.91	22.18 ± 1.24 ^{bE}	14.56 ± 0.44 ^b
0.25	130.40 ± 3.14	26.74 ± 0.65 ^{aD}	16.41 ± 0.11 ^a
0.5	135.07 ± 5.48	29.77 ± 2.81 ^{aC}	17.08 ± 0.19 ^a
1.0	132.97 ± 4.32	28.09 ± 1.55 ^{aCD}	16.72 ± 0.96 ^a
Two-way ANOVA			
Stress	P < 0.001 De	P < 0.001 De	P < 0.001 De
Diet	NS	P < 0.001	P < 0.001
Interaction	NS	P = 0.002	NS

329

330

331

332 4. Discussion

333 Pesticide-contaminated water is a major globally widespread problem in land-based aquaculture
334 [14,51]. Both direct pesticide application in integrated agriculture-aquaculture systems or indirect soil
335 leakage can result in fish being exposed to sub-lethal concentrations of pesticides, which have serious
336 implications for the fish health [14,20,22]. In this study, we have evaluated whether diet
337 supplementation with black seed could mitigate or lower the toxic effects of glyphosate herbicide in
338 common carp fingerlings. Our results show that fish fed with the plant-supplemented diets coped
339 better with glyphosate exposure than control fish, as shown by more stable levels of biochemical
340 serum parameters (total protein, albumin, triglycerides, LDL, cholesterol and HDL), lower levels of
341 metabolic enzymes (AST, ALT, ALP), cortisol and lipid peroxidation (MDA) and higher levels of
342 antioxidant enzymes (SOP and GPx), lysozyme and immunoglobulin than control fish.

343 The immune depression effect of glyphosate on aquatic species such as the Chinese mitten crab,
344 tilapia and silver catfish has previously been observed [22,24,52,53]. Here we show sub-lethal
345 glyphosate exposure for 14 days induced a decrease in the levels of lysozyme and immunoglobulin in
346 common carp fingerlings. Lysozyme is a proteolytic enzyme playing an important role in the innate
347 immune system by killing pathogenic bacteria and triggering other immune responses such as the
348 complement system and phagocytic cells (Saurabh and Sahoo, 2008). Immunoglobulins are also
349 involved in immunity (innate and adaptative) by producing specific antibodies against various
350 antigens [54]. Therefore, decrease in immune responses such as lysozyme and immunoglobulins can
351 have severe impacts in fish health, by lowering their disease resistance. In fact, increased fish disease
352 risk and associated mortality has been observed in fish (*Galaxias anomalus*) exposed to glyphosate
353 [25]. Since, the use of plant-supplements, including the plant species studied here, *N. sativa*, are
354 known to increase fish immune responses [39,55], they arise as an interesting alternative to mitigate at
355 least some of the side-effects of glyphosate exposure in fish. In fact, here we show that common carp
356 supplemented with different levels of *N. sativa*, displayed significantly higher levels of lysozyme,
357 immunoglobulin and total protein (often regarded as an indication of the quantity of immune-related

358 proteins) than fish fed with a basal diet, highlighting the bio-remediation potential of plant-
359 supplemented diets in fish exposed to pesticides such as glyphosate.

360 Several studies have shown that glyphosate exposure in aquatic animals induced oxidative stress,
361 which if maintained for long periods can cause oxidative cell damage [24,53]. Pesticide-related
362 oxidative stress can be induced by the generation of reactive oxygen species (ROS) or by directly
363 interacting with the lipid membranes. In fact, one of the main mechanisms of organophosphate
364 pesticide toxicity is through their interaction with the cytoplasmic membrane [56]. Here we used
365 MDA, a molecule resulting from oxidation processes and known to display toxic effects, as an
366 indicator of lipid peroxidation [57]. Our results show that MDA significantly increased in fish
367 exposed to glyphosate as previously observed in the Chinese mitten crab [24], but fish fed with diets
368 containing *N. sativa* displayed significantly lower levels of MDA. Antioxidant enzymes such as CAT,
369 GPx and SOD have protective effects by preventing uncontrolled generation of ROS and can be an
370 important adaptation to pollutant-induced stress [58]. Previous studies have shown that pesticides
371 including glyphosate inhibited fish antioxidant defences [51,53,58]. Here, levels of SOD and GPx of
372 fish fed with a basal diet were slightly lower after glyphosate exposure, but the decrease was not
373 statistically significant. However, fish fed with supplemented black seed diets displayed significantly
374 higher values, suggesting that inclusion of *N. sativa*, could have activated some antioxidant enzymes
375 (SOD and GPx), which could be related to the lower oxidative stress levels (i.e. MDA) observed in
376 the treated fish.

377 Changes and dysfunction of fish metabolism and biochemical processes have been observed after
378 animal exposure to toxic substances such as pesticides [59]. Therefore, changes in some biochemical
379 parameters such as metabolic enzymes or biomarker molecules of certain metabolic paths (e.g. lipid,
380 carbohydrate, protein) have been proposed as diagnostic tools in toxicology to assess the fish health
381 status and identify the extent of damage to target organs [60–62]. Aminotransferases (ALT, AST) are
382 involved in the metabolism of amino acids, whilst ALP is a polyfunctional enzyme involved in
383 membrane transport activities [61,63]. Our results are in accordance with previous literature showing
384 a significant increase in ALT, AST and ALP in fish exposed to pesticides, including glyphosate

385 [59,64,65]. The increase levels of these enzymes in blood serum was probably related to cytolysis and
386 enzymes leakage into the blood stream, indicating tissue damage in organs such as the liver and
387 kidney [64]. Our results also show, however, that fish fed with diets supplemented with black seed
388 displayed significantly lower levels of ALT, AST and ALP after glyphosate exposure in a dose-
389 dependent manner, highlighting the protective potential of *N. sativa* supplementation on the
390 metabolism of common carp fingerlings, which might be related to significantly lower stress levels
391 (i.e. cortisol). We also observed that glyphosate exposure affected the fish lipid metabolism, by
392 inducing significant reductions in total cholesterol, triglycerides, LDL and HDL. Similar results were
393 observed in tilapia (*Oreochromis niloticus*) exposed to glyphosate for 80 days. Feeding fish with an
394 enriched diet in *N. sativa*, mitigated partly these effects, since fish fed with the treated diet
395 experimented smaller decreases in these parameters and overall, significantly higher levels at the end
396 of the 14-day glyphosate exposure.

397 Finally, the inclusion of *N. sativa* in diets also resulted in increased *C. carpio* fingerlings growth,
398 feeding efficiency and improved digestive enzyme activities. Since pesticides are known to alter and
399 decrease digestive enzymes [66], *N. sativa* supplementation could also display a protective effect on
400 this regard. However, this was not evaluated in this study and needs to be further investigated.

401 In summary, we have shown that sub-lethal exposure of common carp fingerling to glyphosate
402 increases oxidative stress, decreases antioxidant defences, affects several metabolic pathways, and
403 induced immune depression. Furthermore, we show that dietary inclusion of black seed can be used as
404 a sustainable bio-remediation strategy, mitigating many of the negative effects of glyphosate
405 exposure. Fish fed with enriched diets displayed higher levels immune defences (lysozyme and
406 immunoglobulin), lower lipid peroxidation (MDA), higher antioxidant enzymes (SOD, GPx), lower
407 metabolic enzymes (ALT, AST and ALP) in blood serum and lower cortisol levels. Although, more
408 studies are needed to elucidate how pesticides and in particular glyphosate might accumulate in fish
409 muscle, which could cause further human health issues (Lazartigues et al. 2013), our study shows the
410 potential of dietary plant supplementation to lower glyphosate-related toxicity in fish, contributing to
411 stable and environmentally friendly fish production.

412 Data Availability Statement

413 Research data are not shared.

414 Acknowledgement

415 There is no conflict of interest about this article. This paper has been supported by the RUDN
416 University Strategic Academic Leadership Program. This research work was partially supported by
417 Chiang Mai University.

418

419 Reference

- 420 [1] UN, The Sustainable Development Goals Report 2018 | Multimedia Library - United Nations
421 Department of Economic and Social Affairs, New York, 2018.
422 [https://www.un.org/development/desa/publications/the-sustainable-development-goals-report-](https://www.un.org/development/desa/publications/the-sustainable-development-goals-report-2018.html)
423 [2018.html](https://www.un.org/development/desa/publications/the-sustainable-development-goals-report-2018.html) (accessed September 15, 2020).
- 424 [2] S.H. Thilsted, A. Thorne-Lyman, P. Webb, J.R. Bogard, R. Subasinghe, M.J. Phillips, E.H.
425 Allison, Sustaining healthy diets: The role of capture fisheries and aquaculture for improving
426 nutrition in the post-2015 era, *Food Policy*. 61 (2016) 126–131.
427 doi:10.1016/j.foodpol.2016.02.005.
- 428 [3] H.E. Froehlich, C.A. Runge, R.R. Gentry, S.D. Gaines, B.S. Halpern, Comparative terrestrial
429 feed and land use of an aquaculture-dominant world, *Proc. Natl. Acad. Sci. U. S. A.* 115
430 (2018) 5295–5300. doi:10.1073/pnas.1801692115.
- 431 [4] O. Carnevali, F. Maradonna, G. Gioacchini, Integrated control of fish metabolism, wellbeing
432 and reproduction: The role of probiotic, *Aquaculture*. 472 (2017) 144–155.
433 doi:10.1016/j.aquaculture.2016.03.037.
- 434 [5] C. Béné, R. Arthur, H. Norbury, E.H. Allison, M. Beveridge, S. Bush, L. Campling, W.
435 Leschen, D. Little, D. Squires, S.H. Thilsted, M. Troell, M. Williams, Contribution of

- 436 Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current
437 Evidence, *World Dev.* 79 (2016) 177–196. doi:10.1016/j.worlddev.2015.11.007.
- 438 [6] B. Belton, S.R. Bush, D.C. Little, Not just for the wealthy: Rethinking farmed fish
439 consumption in the Global South, *Glob. Food Sec.* 16 (2018) 85–92.
440 doi:10.1016/j.gfs.2017.10.005.
- 441 [7] R. Rufchaei, S.H. Hoseinifar, S. Nedaei, T. Bagheri, G. Ashouri, H. Van Doan, Non-specific
442 immune responses, stress resistance and growth performance of Caspian roach (*Rutilus*
443 *caspius*) fed diet supplemented with earthworm (*Eisenia foetida*) extract, *Aquaculture.* 511
444 (2019) 734275. doi:10.1016/j.aquaculture.2019.734275.
- 445 [8] WorldBank, FISH TO 2030 Prospects for Fisheries and Aquaculture WORLD BANK
446 REPORT NUMBER 83177-GLB, Washington, DC, 2013. www.worldbank.org (accessed
447 September 18, 2020).
- 448 [9] M.G. Bondad-Reantaso, R.P. Subasinghe, J.R. Arthur, K. Ogawa, S. Chinabut, R. Adlard, Z.
449 Tan, M. Shariff, Disease and health management in Asian aquaculture, *Vet. Parasitol.* 132
450 (2005) 249–272. doi:10.1016/j.vetpar.2005.07.005.
- 451 [10] F.C. Cabello, H.P. Godfrey, A.H. Buschmann, H.J. Dölz, Aquaculture as yet another
452 environmental gateway to the development and globalisation of antimicrobial resistance,
453 *Lancet Infect. Dis.* 16 (2016) e127–e133. doi:10.1016/S1473-3099(16)00100-6.
- 454 [11] N. Ahmed, S.W. Bunting, S. Rahman, C.J. Garforth, Community-based climate change
455 adaptation strategies for integrated prawn-fish-rice farming in Bangladesh to promote social-
456 ecological resilience, *Rev. Aquac.* 6 (2014) 20–35. doi:10.1111/raq.12022.
- 457 [12] S.D. Shifflett, A. Culbreth, D. Hazel, H. Daniels, E.G. Nichols, Coupling aquaculture with
458 forest plantations for food, energy, and water resiliency, *Sci. Total Environ.* 571 (2016) 1262–
459 1270. doi:10.1016/j.scitotenv.2016.07.161.

- 460 [13] A.D. Zajdband, Integrated Agri-Aquaculture Systems. In: Lichtfouse E. (eds) Genetics,
461 Biofuels and Local Farming Systems. Sustainable Agriculture Reviews, in: Springer,
462 Dordrecht, 2011: pp. 87–127. doi:10.1007/978-94-007-1521-9_4.
- 463 [14] A. Lazartigues, M. Thomas, C. Cren-Olivé, J. Brun-Bellut, Y. Le Roux, D. Banas, C. Feidt,
464 Pesticide pressure and fish farming in barrage pond in Northeastern France. Part II: Residues
465 of 13 pesticides in water, sediments, edible fish and their relationships, Environ. Sci. Pollut.
466 Res. 20 (2013) 117–125. doi:10.1007/s11356-012-1167-7.
- 467 [15] K.A. Sumon, A. Rico, M.M.S. Ter Horst, P.J. Van den Brink, M.M. Haque, H. Rashid, Risk
468 assessment of pesticides used in rice-prawn concurrent systems in Bangladesh, Sci. Total
469 Environ. 568 (2016) 498–506. doi:10.1016/j.scitotenv.2016.06.014.
- 470 [16] S. Ullah, M.J. Zorriehzahra, Ecotoxicology: A Review of Pesticides Induced Toxicity in Fish,
471 Adv. Anim. Vet. Sci. 3 (2015) 40–57. doi:10.14737/journal.aavs/2015/3.1.40.57.
- 472 [17] S. Spycher, S. Mangold, T. Doppler, M. Junghans, I. Wittmer, C. Stamm, H. Singer, Pesticide
473 Risks in Small Streams - How to Get as Close as Possible to the Stress Imposed on Aquatic
474 Organisms, Environ. Sci. Technol. 52 (2018) 4526–4535. doi:10.1021/acs.est.8b00077.
- 475 [18] H. Rajabi Islami, Y. Filizadeh, Toxicity determination of three sturgeon species exposed to
476 glyphosate, Iran. J. Fish. Sci. 10 (2011) 383–392. <http://jifro.ir/article-1-211-en.html> (accessed
477 July 7, 2020).
- 478 [19] D.W. Kolpin, E.M. Thurman, E.A. Lee, M.T. Meyer, E.T. Furlong, S.T. Glassmeyer, Urban
479 contributions of glyphosate and its degradate AMPA to streams in the United States, Sci. Total
480 Environ. 354 (2006) 191–197. doi:10.1016/j.scitotenv.2005.01.028.
- 481 [20] N.C. Moreno, S.H. Sofia, C.B.R. Martinez, Genotoxic effects of the herbicide Roundup
482 Transorb® and its active ingredient glyphosate on the fish *Prochilodus lineatus*, Environ.
483 Toxicol. Pharmacol. 37 (2014) 448–454. doi:10.1016/j.etap.2013.12.012.

- 484 [21] F.R. de Moura, K.R. Brentegani, A. Gemelli, A.P. Sinhoin, V.D.G. Sinhoin, Oxidative stress
485 in the hybrid fish jundiara (*Leiarius marmoratus* × *Pseudoplatystoma reticulatum*) exposed to
486 Roundup Original®, Chemosphere. 185 (2017) 445–451.
487 doi:10.1016/j.chemosphere.2017.07.030.
- 488 [22] Y. Hong, X. Yang, G. Yan, Y. Huang, F. Zuo, Y. Shen, Y. Ding, Y. Cheng, Effects of
489 glyphosate on immune responses and haemocyte DNA damage of Chinese mitten crab,
490 *Eriocheir sinensis*, Fish Shellfish Immunol. 71 (2017) 19–27. doi:10.1016/j.fsi.2017.09.062.
- 491 [23] A. Topal, M. Atamanalp, A. Uçar, E. Oruç, E.M. Kocaman, E. Sulukan, F. Akdemir, Ş.
492 Beydemir, N. Kiliç, O. Erdoğan, S.B. Ceyhun, Effects of glyphosate on juvenile rainbow
493 trout (*Oncorhynchus mykiss*): Transcriptional and enzymatic analyses of antioxidant defence
494 system, histopathological liver damage and swimming performance, Ecotoxicol. Environ. Saf.
495 111 (2015) 206–214. doi:10.1016/j.ecoenv.2014.09.027.
- 496 [24] X. Yang, Y. Song, C. Zhang, Y. Pang, X. Song, M. Wu, Y. Cheng, Effects of the glyphosate-
497 based herbicide roundup on the survival, immune response, digestive activities and gut
498 microbiota of the Chinese mitten crab, *Eriocheir sinensis*, Aquat. Toxicol. 214 (2019) 105243.
499 doi:10.1016/j.aquatox.2019.105243.
- 500 [25] D.W. Kelly, R. Poulin, D.M. Tompkins, C.R. Townsend, Synergistic effects of glyphosate
501 formulation and parasite infection on fish malformations and survival, J. Appl. Ecol. 47 (2010)
502 498–504. doi:10.1111/j.1365-2664.2010.01791.x.
- 503 [26] E. Awad, A. Awaad, Role of medicinal plants on growth performance and immune status in
504 fish, Fish Shellfish Immunol. 67 (2017) 40–54. doi:10.1016/J.FSI.2017.05.034.
- 505 [27] M. Reverter, N. Tapissier-Bontemps, S. Sarter, P. Sasal, D. Caruso, Moving towards more
506 sustainable aquaculture practices: a meta-analysis on the potential of plant-enriched diets to
507 improve fish growth, immunity and disease resistance, Rev. Aquac. (2020) raq.12485.
508 doi:10.1111/raq.12485.

- 509 [28] S.M. Hoseini, A. Taheri Mirghaed, B.A. Paray, S.H. Hoseinifar, H. Van Doan, Effects of
510 dietary menthol on growth performance and antioxidant, immunological and biochemical
511 responses of rainbow trout (*Oncorhynchus mykiss*), *Aquaculture*. 524 (2020) 735260.
512 doi:10.1016/j.aquaculture.2020.735260.
- 513 [29] E. Ahmadifar, M. Yousefi, M. Karimi, R. Fadaei Raieni, M. Dadar, S. Yilmaz, M.A.O.
514 Dawood, H.M.R. Abdel-Latif, Benefits of Dietary Polyphenols and Polyphenol-Rich Additives
515 to Aquatic Animal Health: An Overview, *Rev. Fish. Sci. Aquac.* (2020).
516 doi:10.1080/23308249.2020.1818689.
- 517 [30] M. Abdel-Tawwab, F. Samir, A.S. Abd El-Naby, M.N. Monier, Antioxidative and
518 immunostimulatory effect of dietary cinnamon nanoparticles on the performance of Nile
519 tilapia, *Oreochromis niloticus* (L.) and its susceptibility to hypoxia stress and *Aeromonas*
520 *hydrophila* infection, *Fish Shellfish Immunol.* 74 (2018) 19–25. doi:10.1016/j.fsi.2017.12.033.
- 521 [31] B.A. Paray, S.M. Hoseini, S.H. Hoseinifar, H. Van Doan, Effects of dietary oak (*Quercus*
522 *castaneifolia*) leaf extract on growth, antioxidant, and immune characteristics and responses to
523 crowding stress in common carp (*Cyprinus carpio*), *Aquaculture*. 524 (2020) 735276.
524 doi:10.1016/j.aquaculture.2020.735276.
- 525 [32] H. Adineh, M. Naderi, A. Nazer, M. Yousefi, E. Ahmadifar, Interactive effects of stocking
526 density and dietary supplementation with Nano selenium and garlic extract on growth, feed
527 utilization, digestive enzymes, stress responses, and antioxidant capacity of grass carp,
528 *Ctenopharyngodon idella*, *J. World Aquac. Soc.* (2020). doi:10.1111/jwas.12747.
- 529 [33] M. Khalili, M. Attar, R. Amirlatifi, Z.N. Maleki, S.M. Hoseini, Effects of dietary myrcene
530 administration on antioxidant gene responses in common carp (*Cyprinus carpio*), exposed to
531 copper sulphate, *Aquac. Res.* 51 (2020) 1653–1659. doi:10.1111/are.14511.
- 532 [34] M. Rabie, Y. Asri, K. Ahmadi, Effect of Milk thistle plant, *Vitis vinifera* extract on immune
533 system of rainbow trout (*Oncorhynchus mykiss*) challenge by diazinon, *Int. J. Aquat. Biol.* 4

- 534 (2016) 208–214. <http://ij-aquaticbiology.com/index.php/ijab/article/view/180> (accessed
535 September 15, 2020).
- 536 [35] S. Hajirezaee, A. Rafieepour, S. Shafiei, R. Rahimi, Immunostimulating effects of Ginkgo
537 biloba extract against toxicity induced by organophosphate pesticide, diazinon in rainbow
538 trout, *Oncorhynchus mykiss*: innate immunity components and immune-related genes, Environ.
539 Sci. Pollut. Res. 26 (2019) 8798–8807. doi:10.1007/s11356-019-04327-7.
- 540 [36] M. Ghelichpour, A. Taheri Mirghaed, Effects of sublethal exposure to new pesticides
541 lufenuron and flonicamid on common carp, *Cyprinus carpio*, hydromineral balance to further
542 saltwater exposure, Int. J. Aquat. Biol. 7 (2019) 195–201. doi:10.22034/ijab.v7i4.662.
- 543 [37] M. Ghelichpour, A. Taheri Mirghaed, S.M. Hoseini, A. Perez Jimenez, Plasma antioxidant and
544 hepatic enzymes activity, thyroid hormones alterations and health status of liver tissue in
545 common carp (*Cyprinus carpio*) exposed to lufenuron, Aquaculture. 516 (2020) 734634.
546 doi:10.1016/j.aquaculture.2019.734634.
- 547 [38] F. Shahid, Z. Farooqui, S. Rizwan, S. Abidi, I. Parwez, F. Khan, Oral administration of *Nigella*
548 *sativa* oil ameliorates the effect of cisplatin on brush border membrane enzymes, carbohydrate
549 metabolism and antioxidant system in rat intestine, Exp. Toxicol. Pathol. 69 (2017) 299–306.
550 doi:10.1016/j.etp.2017.02.001.
- 551 [39] E. Awad, D. Austin, A.R. Lyndon, Effect of black cumin seed oil (*Nigella sativa*) and nettle
552 extract (Quercetin) on enhancement of immunity in rainbow trout, *Oncorhynchus mykiss*
553 (Walbaum), Aquaculture. 388–391 (2013) 193–197. doi:10.1016/j.aquaculture.2013.01.008.
- 554 [40] M. Raymond, Log-probit analysis program presentation for a microcomputer, Entomol.
555 Médicale Parasitol. 23 (1985) 117–121. [https://agris.fao.org/agris-](https://agris.fao.org/agris-search/search.do?recordID=US201302026981)
556 [search/search.do?recordID=US201302026981](https://agris.fao.org/agris-search/search.do?recordID=US201302026981) (accessed August 28, 2020).
- 557 [41] A.E. Ellis, Lysozyme assays, in: J.S. Stolen (Ed.), Techniques in Fish Immunology, in: Fair
558 Haven: SOS publication, 1990: pp. 101–103.

- 559 [42] E. Abdy, M. Alishahi, M. Tollabi, M. Ghorbanpour, T. Mohammadian, Comparative effects of
560 Aloe vera gel and Freund's adjuvant in vaccination of common carp (*Cyprinus carpio* L.)
561 against *Aeromonas hydrophila*, *Aquac. Int.* 25 (2017) 727–742. doi:10.1007/s10499-016-
562 0074-1.
- 563 [43] T. Yano, Assays of hemolytic complement activity, in: J.S. Stolen (Ed.), *Techniques in Fish*
564 *Immunology*, in: Fair haven: SOS publication, 1992: pp. 131–141.
- 565 [44] A. Siwicki, D. Anderson, Nonspecific defense mechanisms assay in fish: II. Potential killing
566 activity of neutrophils and macrophages, lysozyme activity in serum and organs and total
567 immunoglobulin level in serum, in: A. Siwicki, D. Anderson, J. Waluga (Eds.), *Fish Disease*
568 *Diagnos*, in: Olsztyn, Poland, 1993: pp. 105–112.
- 569 [45] S.M. Hoseini, S.H. Hoseinifar, H. Van Doan, Growth performance and hematological and
570 antioxidant characteristics of rainbow trout, *Oncorhynchus mykiss*, fed diets supplemented
571 with Roselle, *Hibiscus sabdariffa*, *Aquaculture*. 530 (2021) 735827.
572 doi:10.1016/j.aquaculture.2020.735827.
- 573 [46] L. Góth, A simple method for determination of serum catalase activity and revision of
574 reference range., *Clin. Chim. Acta.* 196 (1991) 143–51.
575 <http://www.ncbi.nlm.nih.gov/pubmed/2029780> (accessed December 18, 2018).
- 576 [47] M. Mazandarani, S.M. Hoseini, M. Dehghani Ghomshani, Effects of linalool on physiological
577 responses of *Cyprinus carpio* (Linnaeus, 1758) and water physico-chemical parameters during
578 transportation, *Aquac. Res.* 48 (2017) 5775–5781. doi:10.1111/are.13400.
- 579 [48] A. Taheri Mirghaed, M. Ghelichpour, S.M. Hoseini, K. Amini, Hemolysis interference in
580 measuring fish plasma biochemical indicators, *Fish Physiol. Biochem.* 43 (2017) 1143–1151.
581 doi:10.1007/s10695-017-0359-y.
- 582 [49] E. Ahmadifar, N. Sheikhzadeh, K. Roshanaei, N. Dargahi, C. Faggio, Can dietary ginger
583 (*Zingiber officinale*) alter biochemical and immunological parameters and gene expression

- 584 related to growth, immunity and antioxidant system in zebrafish (*Danio rerio*)?, *Aquaculture*.
585 507 (2019) 341–348. doi:10.1016/J.AQUACULTURE.2019.04.049.
- 586 [50] A. Zamani, A. Hajimoradloo, R. Madani, M. Farhangi, Assessment of digestive enzymes
587 activity during the fry development of the endangered Caspian brown trout *Salmo caspius*, *J.*
588 *Fish Biol.* 75 (2009) 932–937. doi:10.1111/j.1095-8649.2009.02348.x.
- 589 [51] A.S. Rossi, N. Fantón, M.P. Michlig, M.R. Repetti, J. Cazenave, Fish inhabiting rice fields:
590 Bioaccumulation, oxidative stress and neurotoxic effects after pesticides application, *Ecol.*
591 *Indic.* 113 (2020) 106186. doi:10.1016/j.ecolind.2020.106186.
- 592 [52] L.. Kreutz, L.. Gil Barcellos, S. de Faria Valle, D. de Oliveira Silva, T Anziliero, E. Davi dos
593 Santos, M. Pivato, R. Zanatta, Altered hematological and immunological parameters in silver
594 catfish (*Rhamdia quelen*) following short term exposure to sublethal concentration of
595 glyphosate, *Fish Shellfish Immunol.* 30 (2011) 51–57. doi:10.1016/j.fsi.2010.09.012.
- 596 [53] T. Zheng, R. Jia, L. Cao, J. Du, Z. Gu, Q. He, P. Xu, G. Yin, Effects of chronic glyphosate
597 exposure on antioxidant status, metabolism and immune response in tilapia (GIFT,
598 *Oreochromis niloticus*), *Comp. Biochem. Physiol. Part - C Toxicol. Pharmacol.* 239 (2021)
599 108878. doi:10.1016/j.cbpc.2020.108878.
- 600 [54] M.R. Coscia, P. Simoniello, S. Giacomelli, U. Oreste, C.M. Motta, Investigation of
601 immunoglobulins in skin of the Antarctic teleost *Trematomus bernacchii*, *Fish Shellfish*
602 *Immunol.* 39 (2014) 206–214. doi:10.1016/j.fsi.2014.04.019.
- 603 [55] Y. Celik Altunoglu, S. Bilen, F. Ulu, G. Biswas, Immune responses to methanolic extract of
604 black cumin (*Nigella sativa*) in rainbow trout (*Oncorhynchus mykiss*), *Fish Shellfish Immunol.*
605 67 (2017) 103–109. doi:10.1016/j.fsi.2017.06.002.
- 606 [56] P. Kavitha, J. Venkateswara Rao, Oxidative stress and locomotor behaviour response as
607 biomarkers for assessing recovery status of mosquito fish, *Gambusia affinis* after lethal effect
608 of an organophosphate pesticide, monocrotophos, *Pestic. Biochem. Physiol.* 87 (2007) 182–

- 609 188. doi:10.1016/j.pestbp.2006.07.008.
- 610 [57] B. Clasen, V.L. Loro, C.R. Murussi, T.L. Tiecher, B. Moraes, R. Zanella, Bioaccumulation
611 and oxidative stress caused by pesticides in *Cyprinus carpio* reared in a rice-fish system, *Sci.*
612 *Total Environ.* 626 (2018) 737–743. doi:10.1016/j.scitotenv.2018.01.154.
- 613 [58] C. Yang, W. Lim, G. Song, Mediation of oxidative stress toxicity induced by pyrethroid
614 pesticides in fish, *Comp. Biochem. Physiol. Part - C Toxicol. Pharmacol.* 234 (2020) 108758.
615 doi:10.1016/j.cbpc.2020.108758.
- 616 [59] P. Samanta, S. Pal, A.K. Mukherjee, A.R. Ghosh, Evaluation of metabolic enzymes in
617 response to Excel Mera 71, a glyphosate-based herbicide, and recovery pattern in freshwater
618 teleostean fishes, *Biomed Res. Int.* 2014 (2014). doi:10.1155/2014/425159.
- 619 [60] A. Van Waarde, M. De Wilde-Van Berge Henegouwen, Nitrogen metabolism in goldfish,
620 *Carassius auratus* (L.). Pathway of aerobic and anaerobic glutamate oxidation in goldfish liver
621 and muscle mitochondria, *Comp. Biochem. Physiol. -- Part B Biochem.* 72 (1982) 133–136.
622 doi:10.1016/0305-0491(82)90021-9.
- 623 [61] S.K. Ramaiah, A toxicologist guide to the diagnostic interpretation of hepatic biochemical
624 parameters, *Food Chem. Toxicol.* 45 (2007) 1551–1557. doi:10.1016/j.fct.2007.06.007.
- 625 [62] S. Falcinelli, A. Rodiles, S. Unniappan, S. Picchietti, G. Gioacchini, D.L. Merrifield, O.
626 Carnevali, Probiotic treatment reduces appetite and glucose level in the zebrafish model, *Sci.*
627 *Rep.* 6 (2016) 1–13. doi:10.1038/srep18061.
- 628 [63] A. Taheri Mirghaed, M. Ghelichpour, A. Zargari, M. Yousefi, Anaesthetic efficacy and
629 biochemical effects of 1,8-cineole in rainbow trout (*Oncorhynchus mykiss*, Walbaum, 1792),
630 *Aquac. Res.* 49 (2018). doi:10.1111/are.13671.
- 631 [64] C. Bacchetta, A. Rossi, A. Ale, M. Campana, M.J. Parma, J. Cazenave, Combined
632 toxicological effects of pesticides: A fish multi-biomarker approach, *Ecol. Indic.* 36 (2014)

- 633 532–538. doi:10.1016/j.ecolind.2013.09.016.
- 634 [65] K.A. Al-Ghanim, S. Mahboob, P. Vijayaraghavan, F.A. Al-Misned, Y.O. Kim, H.J. Kim, Sub-
635 lethal effect of synthetic pyrethroid pesticide on metabolic enzymes and protein profile of non-
636 target Zebra fish, *Danio rerio*, Saudi J. Biol. Sci. 27 (2020) 441–447.
637 doi:10.1016/j.sjbs.2019.11.005.
- 638 [66] A.K. Gupta, P. Pandey, S. Srivastava, Effect of pesticides on the enzyme activity in the fish,
639 *Channa striatus*, Plant Arch. 7 (2007) 749–751.

640

641

642

Highlights

- Glyphosate induced oxidative stress (MDA) and immune suppression in common carp.
- Dietary supplementation with black seed lowered glyphosate toxicity in common carp.
- Carps fed with black seed had higher levels of immune defences than control fish.
- Carps fed with black seed had higher antioxidant enzymes than control fish.

Journal Pre-proof